Three-dimensional conformal intensity-modulated radiation therapy of left femur foci does not damage the sciatic nerve

Wanlong Xu1, Xibin Zhao1, Qing Wang1, Jundang Sun1, Jiangbo Xu1, Wenzheng Zhou1, Hao Wang1, Shigui Yan2, Hong Yuan1

1 People’s Hospital of Xinjiang Uygur Autonomous Region, Urumqi, Xinjiang Uygur Autonomous Region, China
2 The Second Affiliated Hospital of Zhejiang University School of Medicine, Hangzhou, Zhejiang Province, China

Abstract

During radiotherapy to kill femoral hydatid tapeworms, the sciatic nerve surrounding the focus can be easily damaged by the treatment. Thus, it is very important to evaluate the effects of radiotherapy on the surrounding nervous tissue. In the present study, we used three-dimensional, conformal, intensity-modulated radiation therapy to treat bilateral femoral hydatid disease in Meriones meridiani. The focus of the hydatid disease on the left femur was subjected to radiotherapy (40 Gy) for 14 days, and the right femur received sham irradiation. Hematoxylin-eosin staining, electron microscopy, and terminal deoxynucleotidyl transferase–dUTP nick end labeling assays on the left femurs showed that the left sciatic nerve cell structure was normal, with no obvious apoptosis after radiation. Trypan blue staining demonstrated that the overall protoscolex structure in bone parasitized with Echinococcus granulosus disappeared in the left femur of the animals after treatment. The mortality of the protoscolex was higher in the left side than in the right side. The succinate dehydrogenase activity in the protoscolex in bone parasitized with Echinococcus granulosus was lower in the left femur than in the right femur. These results suggest that three-dimensional conformal intensity-modulated radiation therapy achieves good therapeutic effects on the secondary bone in hydatid disease in Meriones meridiani without damaging the morphology or function of the sciatic nerve.

Key Words: nerve regeneration; three-dimensional conformal intensity-modulated radiation therapy; hydatid disease; sciatic nerve; neurons; radiation damage; succinate dehydrogenase; NSFC grant; neural regeneration

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Introduction

Hydatid disease, also called echinococcosis, is a parasitic disease of Echinococcus tapeworms, which is a zoonotic infection in humans (Wang et al., 2013). Bone hydatid disease occurs when echninococcus larvae parasitizes bone tissue. Recently, three-dimensional conformal intensity-modulated radiation therapy has been frequently used clinically to treat tumors because of its advantages, including efficacy in killing tumors and protection of the peripheral organs. Conformal intensity-modulated radiation therapy has been demonstrated to kill bone hydatid cysts without injuring the surrounding tissue (Xu et al., 2012a, b, d). However, because the anatomical structure of the femur is complex, it remains unclear whether conformal intensity-modulated radiation therapy would harm the sciatic nerve. The aim of the present study was to assess the efficacy of conformal intensity-modulated radiation therapy on killing hydatid cysts in the femur of Meriones meridiani and whether it damages the sciatic nerve, in order to determine the efficacy and safety of conformal intensity-modulated radiation therapy in the treatment of hydatid disease in bone.

Materials and Methods

Establishment of animal models of hydatid disease in bone

One hundred and twenty purebred, clean, adult, male Meriones meridiani aged 2–3 months and weighing 30.0 ± 6.5 g were provided by the Institute of Experimental Animals, Chinese Armed Police General Hospital (license No. SCXK (Jing) 2011-0012). The animals were housed in individual cages, fed with complete feed and clean water at 20–23°C and humidity of 40–60%, and ensured adequate sleep. The experimental protocol was approved by the Animal
Ethics Committee of the People’s Hospital of Xinjiang Uyghur Autonomous Region in China (Ethical approval No. 20130301A41). The superiosteum of the hind femurs of Meriones meridiani was selected as the best position for hydatid inoculation with a suspension of 12,000 scolex/mL. The hydatid suspension was injected into the superiosteum of the hind femur of Meriones meridiani using a No. 7 needle. Three months later, X-ray and CT examinations verified a successful inoculation of secondary hydatid in the femur.

Radiotherapy
The regions in the 120 Meriones meridiani with hydatid cysts were considered to be the target areas. A successful treatment strategy would not harm the surrounding blood vessels, tendons, nerves, and skin. CT scanning (Philips, Cleveland, OH, USA) and computer-aided design were used to outline the target areas. After formulating a radiotherapy plan, the Meriones meridiani received inhalation anesthesia and euthanized by air embolism. Un

Sample collection
After conformal intensity-modulated radiation therapy, the Meriones meridiani with hydatid disease were subjected to inhalation anesthesia and euthanized by air embolism. Under an operating microscope (Sichuan Keaoa Company, The Institute of Optics and Electronics, The Chinese Academy of Sciences, Chengdu, Sichuan Province, China), cysts from the Echinococcus granulosus were harvested from the femoral marrow cavities bilaterally. Portions of the hydatid fluid were aspirated from the cysts to stabilize intracapsular pressure. Then, the remaining protoscoleces of the Echinococcus granulosus and hydatid fluid were aspirated. All the obtained samples were mixed and made into suspensions, which were separately placed in sterile polyethylene tubes. Bilateral sections of the sciatic nerve after radiotherapy were obtained, rapidly fixed in 10% formaldehyde and 2.5% glutaraldehyde, and stored in liquid nitrogen. After paraffin imbedding, all samples were sliced into 4 μm-thick sections, dewaxed, hydrated, and stained with hematoxylin and eosin. The histological morphology of the sciatic nerves was observed under light microscopy (Leica, Wetzlar, Germany).

Trypan blue staining to detect pathological changes in the bone Echinococcus granulosus and the protoscoleces mortality after radiation therapy
The suspensions of protoscoleces from the Echinococcus granulosus in sterile polyethylene tubes were stained with 1% trypan blue solution (Santa Cruz Biotechnology, Santa Cruz, CA, USA) for approximately 2 minutes and then smeared on the slides. The percentage of necrotic protoscolex was judged using visual fields on an inverted microscope (Leica). Dead protoscoleces show damaged structure and are stained with trypan blue, while live scolex have clear structure and are not stained with trypan blue. The percentage of necrotic cells was calculated as the number of protoscolex stained with trypan blue/the total number of protoscolex × 100%.

Enzyme histochemical staining to detect succinate dehydrogenase activity in protoscoleces
The smears of the suspensions of Echinococcus granulosus protoscoleces in the sterile polyethylene tubes were dried with a hair dryer using the nitroblue tetrazolium (Santa Cruz Biotechnology) method. Then, the samples were incubated in incubation solution at 25–30°C for 50 minutes until the blue color had fully developed. After being washed in distilled water, all the samples were fixed in 10% calcium formate for 10 minutes, washed with running water for 3 minutes, and then observed with a microscope (Leica). Using the BT-2000 color pathology image analysis system (Hubei Botai Electronic Technology Co., Ltd., Wuhan, Hubei Province, China), 10 fields were randomly selected from each image and mean optical density was measured.

Ultrastructure of the sciatic nerve by transmission electron microscopy
After washing with buffer solution, the samples were pre-fixed in 2.5% glutaraldehyde for 2 hours, post-fixed in 1% osmic acid for 2 hours, and then washed with PBS. The samples were dehydrated with increasing concentrations of ethanol and acetone, embedded, dried at 37°C overnight, sliced into 50 nm-thick sections, and observed and imaged with a transmission electron microscope (Hitachi, Hokkaido, Japan).

Apoptosis of sciatic nerve neurons detected by terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) after radiotherapy
Paraffin sections of the sciatic nerves were washed twice with xylene for 4–5 minutes, rehydrated through a graded alcohol series in 1 minute intervals, incubated in protease at 37°C for 20–30 minutes, incubated in transparent liquid for 6 minutes, and washed twice with PBS. When the slides were dry, the sections were incubated with 50 μL of TUNEL reagent (Boehringer Mannheim, Berlin, Germany) at 37°C for 1 hour, and then were washed three times with PBS. After the slides were dry, the sections were incubated with POD (Boehringer Mannheim) at 37°C for 30 minutes, washed three times with PBS, incubated with 100 μL of diamobenzidine at 15–25°C for 10 minutes, washed three times with PBS, and then stained with hematoxylin and diamobenzidine (Boehringer Mannheim). Next, the slides were dehydrated through a graded alcohol series, permeabilized with xylene, and mounted with a neutral resin. Using the BT-2000 color pathology image analysis system (Hubei Botai Electronic Technology Co., Ltd.), 10 fields at 20 × magnification with densely distributed apoptotic cells were selected.
from each section. The neuronal apoptotic index (%) was calculated as the mean optical density of the TUNEL-positive cells × the percentage of TUNEL-positive cells (from all types of cells in the field) (Stock et al., 2012).

Statistical analysis
Data were analyzed using SPSS 19.0 software (IBM, Armonk, NY, USA) and expressed as the mean ± SD. The group means were compared using Student’s t-tests (α = 0.05). Values of P < 0.05 were considered statistically significant.

Results
Conformal intensity-modulated radiation therapy did not alter the morphology of sciatic nerve neurons in the femur of Meriones meridiani with secondary hydatid disease
Hematoxylin–eosin staining showed a normal morphology of the sciatic nerve neurons in the left femur of Meriones meridiani with osseous hydatid disease after radiotherapy. Although a few neuronal cells showed reversible changes, most cells were triangular or polygonal with clear dendrites and axons. The nuclei were plump, and the nuclear structure was normal. Abundant Nissl’s bodies were uniformly distributed in the cytoplasm, and no clear differences were found between the sciatic nerve neurons in the right side that did not receive radiotherapy (Figure 1).

The transmission electron microscopy results demonstrated that the sciatic nerve neurons in the left side of Meriones meridiani with osseous hydatid disease were normal after conformal intensity-modulated radiation therapy. The cell membranes were not ruptured, the organelles in the cytoplasm were complete, and the nuclei were in the center, with no visible pyknosis. No significant differences were found compared to the neurons in the right side that did not receive radiotherapy (Figure 2).

Conformal intensity-modulated radiation therapy increased the percentage of dead protoscolex in the bone Echinococcus granulosus
In the right femur, which did not receive radiotherapy, the shape of the protoscolex of Echinococcus granulosus was normal in the region with echinococcosis granulosa, and the edge was distinct, repelling trypan blue and remaining unstained. However, after radiotherapy, the protoscolex in the left femur disappeared in the region with Echinococcus granulosus, the cell edges were not distinct, and the cells stained positively for trypan blue. The percent age of dead protoscolex in the osseous hydatid was increased in the region with echinococcosis granulosa after radiotherapy (P < 0.05; Figure 3).
Conformal intensity-modulated radiation therapy decreased succinate dehydrogenase activities in protoscolex

Enzyme histochemical staining showed a high activity of succinate dehydrogenase in the protoscolex of the osseous hydatid in the right femur, which did not receive radiotherapy. The succinate dehydrogenase activity was decreased by radiotherapy in the left femur. The mean optical density of the protoscolex in the osseous hydatid was lower in the left femur than in the right femur ($P < 0.05$; Figure 4).

Conformal intensity-modulated radiation therapy did not alter the apoptosis index of sciatic nerve neurons in Meriones meridiani with secondary hydatid disease

The apoptotic index, calculated from the TUNEL staining, was not different between the left and right femurs in the animals ($P > 0.05$; Figure 5).

Discussion

Since the turn of the century, the rapid progress in imaging and computer technologies has created an era of precision radiotherapy. Of the different radiotherapy techniques, three-dimensional conformal intensity-modulated radiation therapy has become the most widely used. Intensity-modulated radiation therapy controls the direction of the radiation using a computer. Precise, high-dose radiation can be focused on the diseased tissue area of concern, while only rarely damaging the surrounding tissue. Intensity-modulated radiation therapy is a reliable, high-precision radiation therapy used in the clinic, which is also highly praised by scholars (Xilinbaoleri et al., 2009; Cho et al., 2010; Hevezi, 2010; Lisbona et al., 2010; Nagesha et al., 2010; Barney et al., 2011; Chen et al., 2011; Li et al., 2011; Peng et al., 2011; Sangalli et al., 2011; Shi et al., 2011; Teke et al., 2011; Valeriani et al., 2011; Graff et al., 2012; Lee et al., 2012; Mohammed et al., 2012; Stock et al., 2012; Bettington et al., 2013; Bonnette, 2013; Ichinohe et al., 2013; Kim et al., 2013; Kobayashi et al., 2013; Li et al., 2013; Margaritora et al., 2013; Perri et al., 2013; Qian et al., 2013; Salama and Vokes, 2013; Sun et al., 2013; Sung et al., 2013; Tao et al., 2013; Yin et al., 2013;
Zhang et al., 2013). Because intensity-modulated radiation therapy is effective at killing tumor cells, we believed that it would be effective for killing the protoscolex of *Echinococcus granulosus*. Thus, in the present study, we performed intensity-modulated radiation therapy, trypan blue staining, and histochemical staining, which showed that intensity-modulated radiation therapy effectively killed the protoscolex of *Echinococcus granulosus* within the cancellous bone and trabeculae and decreased the residual and relapse rates of bone hydatid.

The results of this study indicated that intensity-modulated radiation therapy decreased the activity of succinate dehydrogenase in the protoscolex of *Echinococcus granulosus*. Succinate dehydrogenase is a rate-limiting enzyme in the Krebs cycle and cell respiration chain, is involved in cellular energy metabolism and oxygen utilization, and is necessary for cell metabolism and vital movement. The succinate dehydrogenase activity reflects the workings of the tricarboxylic acid cycle, and indirectly reflects the levels of cellular aerobic glucose metabolism (Xu et al., 2014). In the present study, intensity-modulated radiation therapy decreased the succinate dehydrogenase activity in the protoscolex of *Echinococcus granulosus*, likely causing a reduction in cellular energy metabolism and eventually cell death. Our experimental results are consistent with those of other papers. Chen and Xie (2014) reported that radiotherapy was effective for treating bone hydatid disease in mice, with different radiation doses leading to different hydatid scolex mortalities, but none of the various radiation doses injured the mice. Xu et al. (2012c) performed intensity-modulated radiation therapy for bone hydatid for 1 month and found that the apoptotic index of the cells in the germinal layer of the hydatid cysts was positively correlated with the dose of radiotherapy. Wang et al. (2009) reported that X-ray irradiation could kill echinococcus isolated from rats, and this lethal effect was dose-dependent. Zhang et al. (2011) demonstrated that an appropriate radiation dose inhibited the growth of alveolar hydatid in rats with hydatid disease. Yuan et al. (2013) showed that *Echinococcus granulosus* protoscolecies were directly killed by γ-ray irradiation in vitro and that the protoscolexes were apoptotic.

Sciatic nerve is the tissue that most needs protection during radiotherapy for femoral lesions. However, the light microscopy, electron microscopy, and TUNEL assay results here indicated that the morphology of sciatic nerve neurons was normal after radiotherapy, suggesting that intensity-modulated radiation therapy during the treatment of osteohydatidosis is relatively safe for the surrounding tissues.

In summary, intensity-modulated radiation therapy was effective for the treatment of secondary osteohydatidosis and safe for the surrounding tissue. However, this was a preliminary study using an animal model. Additional clinical studies should be performed before applying the above results in the clinic.

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**Author contributions:** Xu WL, provided data, ensured the integrity of the data and wrote the manuscript. Yuan H participated in study concept and design, and obtained the funding. Zhao XB was in charge of manuscript authorization. Sun JG, Xu JB, Zhou WZ and Wang H provided technical or data support, and served as the principle investigators. All authors approved the final version of the paper.

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