Susceptibility weighted imaging in the evaluation of hemorrhagic diffuse axonal injury

Diffuse axonal injury (DAI) is axonal and small vessel injury produced by a sudden acceleration of the head by an external force, and is a major cause of death and severe disability (Paterakis et al., 2000). Prognosis is poorer in patients with apparent hemorrhage than in those without (Paterakis et al., 2000). Therefore, it is important to identify the presence and precise position of hemorrhagic foci for a more accurate diagnosis. CT and magnetic resonance imaging (MRI) have long been applied in the diagnosis of DAI, but they are not sensitive enough for the detection of small hemorrhagic foci, and cannot meet the requirements for early diagnosis. A major advance in MRI has been the development of susceptibility weighted imaging (SWI), which has greatly increased the ability to detect small hemorrhagic foci after DAI (Ashwal et al., 2006). In this study, we retrospectively analyzed MRI data for 25 patients with hemorrhagic DAI verified by clinical imaging, and we explored whether SWI was sufficiently sensitive to evaluate hemorrhagic DAI and help accurately assess the severity of the disease.

We recruited 25 patients with hemorrhagic DAI verified by clinical imaging from the Wuxi No.2 People’s Hospital Affiliated to Nanjing Medical University in China from December 2007 to June 2014. Inclusion criteria: (1) history of head trauma; (2) coma or protracted unconsciousness immediately after injury; (3) no clear signs of nervous system localization; (4) unclear changes by cranial CT, severe clinical manifestation; (5) MRI reveals hemorrhagic foci (usually, diameter ≤ 20 mm) in the corpus callosum, brain stem, corticomedullary junction or cerebellum; (6) the disease allows MRI examination.

Exclusion criteria: (1) history of brain injury; (2) malformations or developmental disorders of the central nervous system; (3) history of cerebral hemorrhage or infarction; (4) presence of other diseases unrelated to the trauma which may affect consciousness; (5) poor cooperation resulting in poor image quality.

The 25 patients comprised 19 males and 6 females, 13–49 years of age (average: 37.7 years). Of these, 18 cases were injured by road accident, 5 cases were injured by falling, and 2 cases by hitting. Patients’ GCS scores were graded in accordance with the Glasgow Coma Scale (GCS) (Feng et al., 2014). The scale is composed of three tests: eye, verbal and motor responses. The sum of the three aspects represents the degree of disturbance of consciousness. The scores range from 3 (deep unconsciousness to 15 (fully awake). Generally, brain injury is classified as follows: mild, GCS 13–15; moderate, GCS 9–12; and severe, GCS < 3–8. Among our patients, five scored 9–15 (mild and moderate) and 20 scored 3–8 (severe). MRI was performed 1 day to 1 week after injury. Restless patients were intravenously given 5–10 mg diazepam before examination. Critically ill patients were monitored using electrocardiogram (ECG) gating and respiratory gating. Transverse scanning of the skull was conducted with a Signa EXCITE 1.5 T HD TwinSpeed MR scanner (GE, USA) and an 8-channel head coil. Scanning setup was as follows: (1) T1WI: repetition time (TR) = 500 ms; echo time (TE) = 9.5 ms; time of inversion = 300 ms; (2) T2WI: TR = 3,500 ms; TE = 92 ms; (3) FLAIR: TR = 9,000 ms; TE = 120 ms; (4) GRE: TR = 1,200 ms; TE = 30 ms; (5) SWI: TR = 300 ms; TE = 40 ms; (6) T1WI (FIR): TR = 1,200 ms; TE = 80 ms; b = 0, 1,000, gradient echo (GRE) T2*WI: TR = 600 ms; TE = 15 ms; slice thickness = 5 mm; and gap, 1 mm. SWI used high-resolution three-dimensional GRE imaging: 42 ms, echo time, 25 ms; slice thickness, 2 mm; no gap scanning; 116 layers; number of excitations, 0.75. Two groups of images were collected,

magnitude images and phase images, which were loaded into the workstation (GE ADW4.3) for post-processing. Minimum intensity projection (minIP) was utilized for reconstruction to obtain a slice thickness of 5 mm and a gap of 1 mm. Finally, SWI images were obtained.

The position and number of hemorrhagic foci were identified by two experienced physicians according to the characteristics of the focal distributions (with no long continuous abnormal low-signal intensity within multiple levels of SWI), in combination with GRE and conventional methods. Inconsistent findings were identified together by the two doctors. According to the classification of DAI (Paterakis et al., 1998), DAI was classified as big hemorrhagic focus (20 mm ≥ diameter > 10 mm) or small hemorrhagic focus (diameter ≤ 10 mm). The difference in the number of hemorrhagic foci detected with the various methods was compared using the k independent samples test. A value of P < 0.05 was considered statistically significant. Numerical data were analyzed using the Chi-square test. Correlation of the number of foci revealed by SWI and the GCS score was analyzed using Spearman’s method.

Some abnormal signals revealed by SWI were also found on T1WI, T2WI, FLAIR, DWI and GRE. Low signal intensity, high signal intensity or mixed signal intensity were detected on MRI and DWI because of different bleeding time. The edge of the focus was blurred and fused into a sheet. We noted foci of low signal intensity on GRE and SWI. The images of most foci of low signal intensity on both GRE and SWI were slightly bigger on SWI than on GRE. The signal intensity on SWI was lower than on GRE (Figure 1A–E).

Manifestations of hemorrhagic foci after DAI on SWI clearly showed the size, boundary and range of hemorrhagic foci. Spot-like, nodular, patchy and funicular low-signal-intensity foci of different sizes were scattered in the predilection site. Isointense and hyperintense signals were seen in the big hemorrhagic focus. On the phase image, the big, big hemorrhagic focus displayed hyperintense signal; spot-like hypointense and hypointense signals on the central plane. The upper and lower planes near the lesions displayed low signal intensity. The small hemorrhagic focus exhibited low signal intensity. The diameter of the hemorrhagic focus was 1–20 mm (Figure 1F).

A total of 632 hemorrhagic foci were detected in 25 patients with DAI. In a single case, the number of hemorrhagic foci was a minimum of 2 and a maximum of 84. The hemorrhagic foci were mostly found in the white matter and corticomedullary junction (436, 68.99%), followed by the basal ganglia (90, 14.24%), corpus callosum (70, 11.08%), cerebellum (22, 3.48%) and brain stem (14, 2.21%).

Comparison of the number and distribution of the hemorrhagic foci as detected by the different scanning methods is provided in Table 1. Significant differences in the number of hemorrhagic foci were found among the various scanning methods (P < 0.01). According to average rank, the highest number of hemorrhagic foci was detected by SWI (average rank: 120.20), followed by GRE (average rank: 85.22), DWI (average rank: 77.08), FLAIR (average rank: 68.62), T2WI (average rank: 58.02) and T1WI (average rank: 43.86). There were significant differences in the number of hemorrhagic foci in the different regions (white matter, corticomedullary junction, basal ganglia and corpus callosum) among the various scanning methods (P < 0.05). Although SWI detected the highest number of hemorrhagic foci, no significant difference in the number of hemorrhagic foci in the brain stem or cerebellum was found among the different scanning methods (P > 0.05).

The number of hemorrhagic foci of different sizes detected by the various methods following DAI is shown in Table 2. No significant difference in the number of big hemorrhagic foci (20 mm ≤ diameter > 10 mm) was observed among the various scanning methods (P > 0.66). However, significant differences in the number of small hemorrhagic foci (diameter ≤ 10 mm) were seen among the various scanning methods (P < 0.01). According to average rank, the highest number of small hemorrhagic foci was detected by SWI (average rank: 121.68), followed by GRE (average rank: 86.28), DWI (average rank: 78.42), FLAIR (average rank: 69.06), T2WI (average rank: 73.40) and T1WI (average rank: 67.02). The number of hemorrhagic foci detected by SWI was significantly negatively correlated with GCS score in the 25 patients (r = -0.46, P < 0.05).
Figure 2

Table 1 Comparison of the number and distribution of hemorrhagic foci in 25 patients with diffuse axonal injury detected by different scanning methods

<table>
<thead>
<tr>
<th>Scanning methods</th>
<th>White matter and corticomedullary junction</th>
<th>Basal ganglia</th>
<th>Corpus callosum</th>
<th>Brain stem</th>
<th>Cerebellum</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1WI</td>
<td>52</td>
<td>14</td>
<td>11</td>
<td>5</td>
<td>4</td>
<td>86</td>
</tr>
<tr>
<td>T2WI</td>
<td>69</td>
<td>14</td>
<td>16</td>
<td>6</td>
<td>6</td>
<td>111</td>
</tr>
<tr>
<td>FLAIR</td>
<td>89</td>
<td>21</td>
<td>19</td>
<td>6</td>
<td>6</td>
<td>141</td>
</tr>
<tr>
<td>DWI</td>
<td>109</td>
<td>22</td>
<td>25</td>
<td>7</td>
<td>8</td>
<td>171</td>
</tr>
<tr>
<td>GRE</td>
<td>133</td>
<td>31</td>
<td>30</td>
<td>9</td>
<td>7</td>
<td>210</td>
</tr>
<tr>
<td>SWI</td>
<td>436</td>
<td>90</td>
<td>70</td>
<td>14</td>
<td>22</td>
<td>632</td>
</tr>
<tr>
<td>$\chi^2$ value</td>
<td>38.416</td>
<td>17.84</td>
<td>13.07</td>
<td>3.88</td>
<td>3.83</td>
<td>46.06</td>
</tr>
<tr>
<td>$P$ value</td>
<td>&lt; 0.01</td>
<td>&lt; 0.01</td>
<td>0.57</td>
<td>0.57</td>
<td>&lt; 0.01</td>
<td></td>
</tr>
</tbody>
</table>

T1WI: T1 weighted image; T2WI: T2 weighted image; FLAIR: fluid-attenuated inversion recovery; DWI: diffusion weighted image; GRE: gradient echo; SWI: susceptibility weighted image.

Table 2 Comparison of the number of hemorrhagic foci of different sizes after diffuse axonal injury detected by the various scanning methods

<table>
<thead>
<tr>
<th>Diameter of the focus</th>
<th>T1WI</th>
<th>T2WI</th>
<th>FLAIR</th>
<th>DWI</th>
<th>GRE</th>
<th>SWI</th>
<th>$\chi^2$ value</th>
<th>$P$ value</th>
</tr>
</thead>
<tbody>
<tr>
<td>&gt; 10 mm</td>
<td>36</td>
<td>37</td>
<td>41</td>
<td>41</td>
<td>56</td>
<td>3.29</td>
<td>0.06</td>
<td>&lt; 0.01</td>
</tr>
<tr>
<td>≤ 10 mm</td>
<td>50</td>
<td>74</td>
<td>100</td>
<td>130</td>
<td>167</td>
<td>576</td>
<td>51.23</td>
<td>&lt; 0.01</td>
</tr>
</tbody>
</table>

T1WI: T1 weighted image; T2WI: T2 weighted image; FLAIR: fluid-attenuated inversion recovery; DWI: diffusion weighted image; GRE: gradient echo; SWI: susceptibility weighted image.

Because of the high sensitivity of SWI to blood metabolites, SWI reveals many hemorrhagic foci after DAL. In the present study, the number of hemorrhagic foci and the amount of bleeding revealed by SWI were respectively 3–6-fold and 2-fold higher than that shown by conventional GRE sequence (Tong et al., 2003, 2004). Our results demonstrate that SWI detects the greatest number of hemorrhagic foci, followed by GRE. T1WI detected the lowest number of hemorrhagic foci. The number of foci detected using SWI was 3.28-fold that detected by conventional GRE sequence, consistent with previous studies.

We also compared the sensitivity of SWI for hemorrhagic foci of different sizes. No significant difference was found in the number of big hemorrhagic foci (20 mm ≥ diameter > 10 mm) among the different scanning methods, but significant differences were seen in the number of small hemorrhagic foci (diameter ≤ 10 mm) among the different scanning methods (SWI detected the highest number of foci). SWI had higher sensitivity for detecting small hemorrhagic foci than other MR sequences. Furthermore, SWI displays the boundaries and extent of the hemorrhagic foci more clearly than a conventional MRI sequence or traditional GRE sequence. Therefore, SWI is a more sensitive and accurate imaging method for identifying small hemorrhagic foci after traumatic brain injury.

There were significant differences in the number of hemorrhagic foci detected by the various scanning methods in the various regions, including the white matter, corticomedullary junction, basal ganglia and corpus callosum. Among the different scanning methods, SWI detected the greatest number of foci. Thus, SWI has obvious advantages for regions with a high number of foci, especially the basal ganglia and corpus callosum. Nevertheless, there was no significant difference in the number of hemorrhagic foci in the brain stem or cerebellum revealed by the various detection methods. This is probably associated with the low number of hemorrhagic foci in the brain stem and cerebellum after injury, as well as the severe magnetic susceptibility artifacts at the skull base-air interface. Moreover, it should be mentioned that the number of cases in this study was limited.

GCS can objectively assess the severity of traumatic brain injury, and is simple to use. In this study, we analyzed the correlation between the number of hemorrhagic foci detected by SWI and the GCS score, and found that the number of hemorrhagic foci was negatively correlated with GCS score. Therefore, SWI can objectively and accurately evaluate the severity of brain injury. A shortcoming of this study is that we did not perform long-term follow-up of the patients.

A number of problems need attention in the detection of hemorrhagic foci using SWI. For example, distinguishing these from small veins and calcified foci, particularly as small veins vertical to the cross section also display oval low signal intensity. Low-signal-intensity foci continuous along numerous levels should be distinguished from microhemorrhage. Calcified and hemorrhagic foci also show low signal intensity in SWI, but calcification is antimagnetic and hemorrhage is paramagnetic. In SWI phase images, paramagnetic substances exhibit a phase increase in the central level and phase reduction in the upper and lower levels, while antimagnetic substances...
Figure 1 Images of the hemorrhagic focus in a 22-year-old male patient with diffuse axonal injury caused by a road traffic accident. (A) T1WE: patchy and funicular low-signal-intensity focus (Δ) in the white matter of the right frontal lobe and corticomedullary junction. (B) T2WE: this focus displays a patchy and funicular high signal intensity (Δ). (C) FLAIR: in addition to the high signal-intensity focus (Δ), we can also detect a spot-like high-signal-intensity focus in the splenium of the right corpus callosum (Δ). (D) DWI: a spot-like high-signal-intensity focus in the splenium of the right corpus callosum can be detected (Δ). (E) T2*WI: spot-like low-signal-intensity foci in the white matter of the right frontal lobe and corticomedullary junction (Δ) and in the splenium and genu of the right corpus callosum are detected (Δ); (F) SWI: in addition to the foci in the splenium and genu of the right corpus callosum, multiple spot-like low-signal-intensity foci are visible in the body of the right corpus callosum. Multiple spot-like and funicular low-signal-intensity foci are observed in the white matter of the bilateral frontal lobes and corticomedullary junction, and in the periventricular white matter as well. T1WE: T1 weighted image; T2WE: T2 weighted image; FLAIR: fluid-attenuated inversion recovery; DWI: diffusion weighted image; SWI: susceptibility weighted image; R: right; L: left.

Figure 2 Scatterplot of the correlation between the number of hemorrhagic foci detected by SWI and the GCS score.

Using Spearman's method, the number of hemorrhagic foci was significantly negatively correlated with GCS score ($r = -0.82, P < 0.01$). SWI: Susceptibility weighted imaging; GCS: Glasgow Coma Scale; SWI: susceptibility weighted image.

Another issue with diagnosis is distinguishing between contusion and laceration injury. Contusion and laceration of brain often damages the cortices of stress points or contrecoup positions, and results in focal or flaky hemorrhages > 20 mm in diameter. These patients have mild clinical symptoms, and a few experience a transient coma, which do not lead to dysfunction or death. DAI frequently occurs in the corticomedullary junction, corpus callosum, basal ganglia, brain stem and cerebellum, and can produce evident dysfunction and death. Therefore, DAI can be distinguished from contusion and laceration of the brain in hemorrhage location, size, shape, distribution and clinical symptoms. A few foci that are difficult to identify may not be included in the tally.

Other limitations of the use of SWI in the clinic are the long scanning time, the numerous intracranial susceptibility artifacts, and overestimates of hemorrhagic foci. Consequently, long-term follow-up is needed for an accurate prognosis.

In summary, SWI is highly sensitive for the detection of hemorrhagic DAI, and can detect a higher number of small hemorrhagic foci than other MR sequences. Furthermore, SWI can accurately reveal foci characteristics and distribution. The apparently negative correlation between the number of hemorrhagic foci revealed by SWI and the GCS score can provide important information for clinical diagnosis and treatment. Therefore, SWI has clinical value in the diagnosis of DAI.

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References